Genomic DNA Preparation from Plant Material

- 1. Picked one leaf from each plant (approximately 1 inch long or two smaller leaves). Put leaf into an appropriately labeled 1.2 mL microdilution stiptube (USA Scientific; Cat # 1212-8000) or 1.5 mL microcentrifuge tube.
- 2. Grind plant tissue well with a stick into a powder, in liquid nitrogen.
- 3. Remove tubes from liquid nitrogen to warm up before adding extraction buffer.
- 4. Add 400 uL extraction buffer (see recipe at bottom of page).
- 5. Stir the sample with the stick and transfer by pouring the sample into a new labeled 1.5 mL tube (not necessary if starting from 1.5 mL microcentrifuge tube.
- 6. Vortex each tube really well for 30 seconds.
- 7. Spin tubes at max speed for 5 minutes.
- 8. Transfer supernatant (~200 uL) to a new tube using a pipet. Do not pour off and avoid the leaf debris.
- Vortex briefly (~5 seconds) and spin tubes again at max speed for 5 minutes.
- Transfer supernatant (~180 uL) to a new tube using a pipet (do not pour off).
- 11. Add an equal amount (180 uL) of isopropanol and vortex well (10 seconds).
- 12. Incubate at room temperature for 10 minutes.
- 13. Spin tubes at max speed for 10 minutes.
- 14. Carefully remove supernatant with a pipette (do not pour off).
- 15. Wash pellet with 1 mL 80% ethanol. Vortex each sample for ~15 seconds. (this can be a stopping point if necessary: place DNA under EtOH in -20°C overnight and resume the following day)
- 16. Spin tubes for 2 minutes at max speed and pour off EtOH.
- 17. Spin again 1 minute and carefully remove all traces of ethanol with a pipette.
- 18. Air dry tubes for 1 hour. Leave a paper towel over the top.
- 19. Add 100uL of H2O and resuspend pellet by pipet and vortexing (disrupt the pellet as much as possible).
- 20. Spin max speed 5 minutes; transfer supernatant (this is the gDNA) to new tube
- 21. Store gDNA at -20°C.

Extraction Buffer Recipe:

REAGENT	[STOCK]	per/500 mL	<u>per/100 mL</u>
200 mM Tris, pH 7.5 250 mM NaCl	1.0 M 5.0 M	100 mL 25 mL	20 mL 5 mL
25 mM EDTA	0.5 M	25 mL	5 mL
0.5 % SDS	10%	25 mL	5 mL
Water		325 mL	65 mL